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(54) Title: OB RECEPTOR ISOFORMS AND NUCLEIC ACIDS ENCODING THEM			
(57) Abstract			
<p>The <i>ob</i> receptor has numerous isoforms resulting from alternative splicing; three novel isoforms, designated c', f, and g are disclosed. The nucleic acids encoding these isoforms are taught. Also part of the invention are vectors containing the nucleic acid encoding the receptors, host cells transformed with these genes, and assays which use the genes or protein isoforms.</p>			

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Furthermore, the recent cloning of the human receptor for the leptin, the ob-receptor (OB-R), reveals that it is transcribed in the hypothalamus (Tartaglia *et al.* 1995, *Cell* 83:1263-1271; Stephens *et al.* 1995, *Nature* 377: 530-532). In addition, a mutation that

5 results in premature termination of the long-form of the mouse OB-R, which is preferentially expressed in the hypothalamus, appears to be responsible for the obese phenotype of the *db/db* mouse (Lee *et al.* 1996, *Nature* 379:632-635; Chua *et al.* 1996, *Science* 271:994-996; and Chen *et al.* 1996, *Cell* 84:491-495).

10 The OB-R from wild type (lean) rats and from rats having the *fatty* mutation (both heterozygous and homozygous *fa*) have been isolated and sequenced. (Patent Application Serial Nos. _____, Attorney Docket Nos. 19642PV and 19642PV2, filed February 22, 1996 and March 22, 1996, which are hereby

15 incorporated by reference.)

Various isoforms of the OB-Rs have also been identified. These isoforms are due to alternative splicing. For example, in the mouse the a form has 5 amino acids following the Lysine at 889; the b form has 273 amino acids after Lysine 889; the c form has 3 amino acids after Lysine 889; and the d form contains 11 amino acids after Lysine 889.

20

It would be desirable to be able to further experiment with various isoforms in order to better understand obesity, and to be able to clone and produce novel *ob* receptor isoforms to use in

25 assays for the identification of ligands which may be useful in understanding obesity and for its prevention and treatment.

DETAILED DESCRIPTION OF THE INVENTION

This invention relates to novel *ob* receptor isoforms

30 designated c', f and g which are substantially free from associated membrane proteins. It also relates to substantially purified *ob* receptor isoform c', f and g proteins. These isoforms are present in various species, including rat, mouse and human.

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Another aspect of this invention is to nucleic acids which encode OB receptor isoforms c', f or g. The nucleic acid may be any nucleic acid which can encode a protein, such as genomic DNA, cDNA, or any of the various forms of RNA. Preferably, the nucleic acid is cDNA.

5 This invention also includes vectors containing a OB-R isoform c', f or g gene, host cells containing the vectors, and methods of making substantially pure OB-R isoform c', f or g protein comprising the steps of introducing a vector comprising a 10 OB-R isoform c', f or g gene into a host cell, and cultivating the host cell under appropriate conditions such that OB-R isoform c', f or g is produced. The OB-R isoform c', f or g so produced may be harvested from the host cells in conventional ways.

15 Yet another aspect of this invention are assays which employ OB-R isoform c', f or g. In these assays, various molecules, suspected of being OB-R isoform c', f or g ligands are contacted with a OB-R isoform c', f or g, and their binding is detected. In this way agonists, antagonists, and ligand mimetics may be identified. A further aspect of this invention are the ligands so identified.

20 **BRIEF DESCRIPTION OF THE FIGURES**

FIGURE 1 is the amino acid sequence of wild type rat OB-R.

25 FIGURE 2 is the cDNA sequence of wild type rat OB-R.
FIGURE 3 is the cDNA sequence encoding rat isoform.
FIGURE 4 is the cDNA specific for Rat isoform c'.

As used throughout the specification and claims, the following definitions apply:

30 "Substantially free from associated membrane proteins" means that the receptor protein is not in physical contact with any membrane proteins.

"Substantially purified OB-receptor isoform c', f or g" means that the protein isoform is at least 90% and preferably at least 95% pure.

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"Wild type" means that the gene or protein is substantially the same as that found in an animal which is not considered to have a mutation for that gene or protein.

5 "fa" means that the gene or protein is substantially the same as that found in a rat homologous for the *fatty* mutation.

"Substantially the same" when referring to a nucleic acid or amino acid sequence means either it is the same as the reference sequence, or if not exactly the same, contains changes which do not affect its biological activity or function.

10

It has been surprisingly found, in accordance with this invention that the OB-R exists in a large variety of isoforms, including three novel ones, form c', f and g. These isoforms apply to all species, but for convenience, throughout the specification and 15 claims, numberings of amino acids and nucleotides will use the rat wild type sequences (FIGURES 1 and 2) as a reference. However, it is to be understood that this invention is not limited to rat wild type proteins and nucleic acids and specifically includes rat (wild type and *fatty*), mouse, and human OB-R isoform c', f and g proteins and 20 nucleic acids.

OB-R isoform f differs from wild type protein in that after the Lysine at position 889 (referring to the rat sequence in FIGURE 1), there are six amino acids, ending at an Asparagine residue at position 895. In the cDNA, the codons are then followed 25 by a Stop codon. One cDNA for rat isoform f is shown in FIGURE 3; this invention specifically includes all various cDNAs encoding an isoform f protein. The superscripted numbers refer to protein position numbers.

30 Lys⁸⁸⁹ Iso⁸⁹⁰ Met⁸⁹¹ Pro⁸⁹² Gly⁸⁹³ Arg⁸⁹⁴ Asn⁸⁹⁵

In the human isoform f, Lysine 891 corresponds to the rat Lysine 889, the same six amino acids follow Lysine 889.

In a particularly preferred embodiment of this invention, the OB-R isoform f is from rat origin.

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OB-R isoform g differs from the wild type in that it is much shorter than the wild type sequence. The following eighteen amino acids are found at the beginning of the protein with the superscript numbers indicating their position. The Arginine at position 18 is spliced to a large fragment of the wild type molecule, beginning at the Proline at position 166 (in both mouse and human). This isoform then extends for the remainder of the wild type molecule.

10 Met¹ Phe² Gln³ Thr⁴ Pro⁵ Arg⁶ Ile⁷ Val⁸ Pro⁹ Gly¹⁰
 His¹¹ Lys¹² Asp¹³ Leu¹⁴ Ile¹⁵ Ser¹⁶ Lys¹⁷ Arg¹⁸ Pro¹⁶⁶...

After Pro¹⁶⁶, the remainder of the protein may be the same as wild type, or, alternatively it could also contain another isoform variation, such as isoform a, b, c, d, e, or f.

15

A particularly preferred embodiment is the rat isoform g.

OB-R isoform c' is similar to the OB-R isoform c which was previously described [Lee *et al.*, *Nature* 379: 632-635]. After Lysine at position 889, it only has three amino acids, Val⁸⁹⁰ Thr⁸⁹¹ Phe⁸⁹² Stop. As can be seen, isoform c' differs from isoform c in that the final amino acid is phenylalanine rather than valine found in isoform c. Further, there are untranslated sequences in the DNA encoding isoform c' which do not appear to be present in isoform c. The cDNA encoding the rat isoform c' is given in FIGURE 4. In humans, the Val, Thr, Phe follow Lysine 891.

30 One aspect of this invention is the molecular cloning of these various isoforms of OB-R. The wild type and *fa* receptor proteins contain an extracellular, a transmembrane domain. In the rat, the extracellular domain extends from amino acids 1-830; the transmembrane domain is from amino acids 839-860; and the cytoplasmic domain is from amino acids 860-1162. Similar domains have been identified for the mouse and human proteins. This

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invention also includes isoform c', f and g proteins which lack one or more of these domains. Such deleted proteins are useful in assays for identifying ligands and their binding activity.

5 In the rat wild type protein, amino acids 1-28 form a signal sequence; thus the mature proteins extend from amino acids 28-1162. The mature protein isoforms form yet another aspect of this invention. This differs somewhat from the signal sequence of 1-22 reported for mouse and human OB-R; the mature mouse and human isoforms form yet another aspect of this invention.

10 The OB-R isoform c', f or g gene can be introduced into virtually any host cell using known vectors. Preferred host cells include *E. coli* as well as mammalian and yeast cell lines.

15 One of ordinary skill in the art is able to choose a known vector which is appropriate for a given host cell; generally plasmids or viral vectors are preferred. The OB-R isoform c', f or g gene may be present in the vector in its native form, or it may be under the control of a heterologous promoter, and if desired, one or more enhancers, or other sequences known to regulate transcription or translation. The host cell containing the OB-R isoform c', f or g gene is cultured, and the OB-R isoform c', f or g gene is expressed. After a suitable period of time the OB-R c', f or g isoform protein may be harvested from the cell using conventional separation techniques.

20 25 A further aspect of this invention is the use of an OB-R c', f or g isoform in assays to identify OB-R c', f or g isoform ligands. A ligand binds to the OB-R isoform receptor, and *in vivo* may or may not result in an activation of the receptor. Ligands may be agonists of the receptor (i.e. stimulate its activity), antagonists (inhibit its activity) or they may bind with little or no effect upon the receptor activity.

30 In an assay for ligands, an OB-R isoform of this invention is exposed to a putative ligand, and the amount of binding is measured. The amount of binding may be measured in many ways; for example, a ligand or the OB-R isoform being investigated

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may be labeled with a conventional label (such as a radioactive or fluorescent label) and then put in contact with the OB-R isoform under binding conditions. After a suitable time, the unbound ligand is separated from the OB-R isoform and the amount of ligand which

5 has bound can be measured. This can be performed with any of the OB-R isoforms of this invention; alternatively the amount of binding of the various isoforms can be compared. In a competitive assay, both the putative ligand and a known ligand are present, and the amount of binding of the putative ligand is compared to the amount

10 of binding to a known ligand. Alternatively, the putative ligand's ability to displace previously bound known ligand (or vice-versa) may be measured. In yet other embodiments, the assay may be a heterogeneous one, where the OB-R isoform may be bound to a surface, and contacted with putative ligands. Detection of binding

15 may be by a variety of methods, including labelling, reaction with antibodies, and chromophores.

In another assay, the OB-R isoforms of this invention may be used in a "trans" activation assay. Such assays are described in U.S. Application Serial No. _____, Attorney Docket No.

20 19686PV, which was filed on April 22, 1996 and which is hereby incorporated by reference. In this assay, a cell which expresses an OB-R isoform of this invention (either naturally or through recombinant means) is transfected with a reporter gene construct comprising a minimal promoter, a leptin activation element and a reporter gene. Transcription of the reporter gene is dependant upon activation of the leptin activation element. Binding of a ligand to the receptor isoform activates the leptin activation element, which then allows transcription of the reporter gene.

25

The following non-limiting Examples are presented to

30 better illustrate the invention.

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EXAMPLE 1

Preparation of mRNA and cDNA from rat tissues

Tissues were collected from lean and *fa/fa* Zucker rats and snap frozen in liquid nitrogen. The tissues collected included: hypothalamus, pituitary, lung, liver, kidney, heart, adrenal glands, smooth muscle, skeletal muscle, and adipose tissue. The tissues were homogenized with a Brinkmann Polytron homogenizer in the presence of guanadinium isothiocyanate. mRNA was prepared from hypothalamus, lung, and kidney according to the instructions provided with the messenger RNA isolation kit (Stratagene, La Jolla, CA). cDNA was prepared from approximately 2 µg of mRNA with the SuperScript™ choice system (Gibco/BRL Gaithersburg, MD). The first strand cDNA synthesis was primed using 1 µg of oligo(dT)12-18 primer and 25 ng of random hexamers per reaction. Second strand cDNA synthesis was performed according to the manufacturer's instructions. The quality of the cDNA was assessed by labeling an aliquot (1/10th) of the second strand reaction with approximately 1 µCi of [α -32P]dCTP (3000 Ci/mmol). The labeled products were separated on an agarose gel and detected by autoradiography.

EXAMPLE 2

Preparation of a hypothalamic cDNA library

Approximately 3.6 µg of phosphorylated *Bst* XI adapters (Invitrogen, San Diego, CA) were ligated to approximately 3 µg of cDNA prepared as described in Example 1. The ligation mix was then diluted and size-fractionated on a cDNA sizing column (Gibco/BRL Gaithersburg, MD). Drops from the column were collected and the eluted volume from the column was determined. An aliquot from each fraction was analyzed on an agarose gel. Fractions containing cDNA of greater than or equal to 1 kb were pooled and precipitated. The size-fractionated cDNA with the *Bst* XI adapters was ligated into the prokaryotic vector pcDNA II

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(Invitrogen, San Diego, CA). The vector (4 μ g) was prepared for ligation by first cutting with the restriction endonuclease *Bst* XI, gel purifying the linearized vector, and then dephosphorylating the ends with calf intestinal phosphatase (Gibco/BRL, Gaithersburg, MD)

5 according to the manufacturers instructions. The ligation contained approximately 10-20 ng of cDNA and approximately 100 ng of vector and was incubated overnight at 14°C. The ligation was transformed into 1 ml of XL-2 Blue Ultracompetent cells (Stratagene, La Jolla, CA) according to the manufacturer's

10 instructions. The transformed cells were spread on 133 mm Colony/Plaque Screen filters (Dupont/NEN, Boston, MA), plated at a density of 30,000 to 60,000 colonies per plate on Luria Broth agar plates containing 100 μ g/ml Ampicillin (Sigma, St. Louis, MO).

15

EXAMPLE 3

Screening a hypothalamic cDNA library

Colonies on filters were replica plated onto a second filter set. The master filter was stored at 4°C for subsequent

20 isolation of regions containing colonies that gave a positive hybridization signal. The replica filters were grown for several hours at 37°C until colonies were visible and then processed for in situ hybridization of colonies according to established procedures (Maniatis, et al. *Molecular Cloning: A Laboratory Manual*, Cold

25 Spring Harbor Laboratory Publications, Cold Spring Harbor, NY, which is hereby incorporated by reference). A Stratalinker (Stratagene, La Jolla, CA) was used to crosslink the DNA to the filter. The filters were washed at 55°C for 2 hours in 2x SSC and 0.5% SDS to remove bacterial debris. Eight to ten filters were then

30 placed in a heat sealable bag (Kapak, Minneapolis, MN) containing 15-20 ml of 1x hybridization solution (Gibco/BRL, Gaithersburg, MD) containing 50% formamide and incubated for 1 hour at 42°C. The filters were hybridized overnight with greater than 1,000,000 cpm/ml of the radiolabeled probe described below in 1x

- 10 -

hybridization buffer (Gibco/BRL, Gaithersburg, MD) containing 50% formamide at 42°C. The probe, a 2.2 kb fragment encoding the extracellular portion of the Ob-R was labeled by random priming with [α 32P]dCTP (3000 Ci/mmol, Amersham, Arlington Heights, IL) using redi-prime (Amersham, Arlington Heights, IL). The probe was purified from unincorporated nucleotides using a Probequant G-50 spin column (Pharmacia Biotech, Piscataway, NJ). Filters were washed two times with 0.1x SSC 0.1% SDS at 60°C for 30 min and then subjected to autoradiography. Individual regions containing hybridization positive colonies were lined up with the autoradiogram of the hybridized filter. These were excised from the master filter, and placed into 0.5 ml Luria broth plus 20% glycerol. Each positive was replated at a density of approximate 50-200 colonies per 100 by 15 mm plate and screened by hybridization as previously described. Individual positive colonies were picked and plasmid DNA was prepared from an overnight culture using a Wizard kit (Promega, Madison, WI).

EXAMPLE 4

20

Amplification of Lean Rat OB-receptor cDNA using PCR

To provide for a probe to screen the hypothalamic cDNA library, the rat OB receptor was initially obtained by PCR using degenerate primers based on the mouse and human OB-receptor amino acid sequences. A set of oligonucleotide primers, were designed to regions with low codon degeneracy. The pairing of the forward primers ROBR 2 (5'-CAY TGG GAR TTY CTI TAY GT-3') and ROBR 3 (5'-GAR TGY TGG ATG AAY GG-3') corresponding to mouse amino acid sequences HWEFLYV and 25 ECWMKG, with reverse primers ROBR 6 (5'-ATC CAC ATI GTR TAI CC-3'), ROBR 7 (5'-CTC CAR TTR CTC CAR TAI CC-3'), ROBR 8 (5'-ACY TTR CTC ATI GGC CA-3') and ROBR 9 (5'-CCA YTT CAT ICC RTC RTC-3') representing mouse amino acids, GYTMWI, VYWSNWS, WPMSKV, and DDGMKW provided good 30 yields of the appropriately sized products. The fragments of interest 35

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were amplified as long polymerase chain reaction (PCR) products by a modifying the method of Barnes (1994, *Proc. Natl. Acad. Sci.* 91:2216-2220, which is hereby incorporated by reference). In order to obtain the required long PCR fragments, Taq Extender

5 (Stratagene, La Jolla CA) and the Expand Long Template PCR System (Boehringer Mannheim, Indianapolis, IN) were used in combination. The standard PCR reaction mix, in a final volume of 20 μ l, contained 5 ng of template (lean rat cDNA), 100 ng of primers, 500 μ M dNTPs, 1 X Buffer 3 from the Expand kit, 0.1 μ l 10 each of Taq Polymerase and Taq Expander. Reactants were assembled in thin walled reaction tubes.

10 The amplification protocol was: 1 cycle of 92°C for 30 sec., followed by 32 cycles at 92°C for 30 sec., 45°C for 1 min. and 68°C for 3 min. using a Perkin-Elmer (Norwalk, CT) 9600 Thermal 15 Cycler.

15 This strategy produced a series of PCR products with the largest being approximately 2.2 Kbp amplified from primers ROBR 2 and ROBR 9. These products were subcloned for DNA sequence analysis as described below. The insert was excised from 20 the cloning vector with the restriction endonuclease *Eco* RI, and fragments were separated from the vector by agarose gel electrophoresis. The fragments were eluted from the gel using a Prep-A-Gene kit (BioRad, Richmond CA) according to the manufacturer's instructions and radiolabeled as described above.

25

EXAMPLE 5

Subcloning of PCR products

30 PCR products of the appropriate size were prepared for subcloning by separation on an agarose gel, excising the band, and extracting the DNA using Prep-A-Gene (BioRad, Richmond, CA). PCR products were ligated into pCRTMII (Invitrogen, San Diego, CA) according to the instructions provided by the manufacturer. The ligation was transformed into INVaF^r cells and plated on Luria- 35 Bertani plates containing 100 μ g/ml ampicillin and X-Gal (32 μ l of

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50 mg/ml X-Gal (Promega, Madison, WI). White colonies were picked and grown overnight in Luria-Bertani broth plus 100 µg/ml ampicillin. Plasmid DNAs were prepared using the Wizard miniprep kit (Promega, Madison, WI). Inserts were analyzed by digesting the 5 plasmid DNA with EcoRI and separating the restriction endonuclease digestion products on an agarose gel.

10 Plasmid DNA was prepared for DNA sequencing by ethanol precipitation of Wizard miniprep plasmid DNA and resuspending in water to achieve a final DNA concentration of 100 µg/ml. DNA sequence analysis was performed using the ABI PRISM™ dye terminator cycle sequencing ready reaction kit with AmpliTaq DNA polymerase, FS. The initial DNA sequence analysis was performed with M13 forward and reverse primers, subsequently 15 primers based on the rat OB-R sequence were utilized. Following amplification in a Perkin-Elmer 9600, the extension products were purified and analyzed on an ABI PRISM 377 automated sequencer (Perkin Elmer, Norwalk, CT). DNA sequence data was analyzed with the Sequencher program.

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WHAT IS CLAIMED IS:

1. *Ob*-receptor (OB-R) isoform c', f or g, substantially free from associated proteins.
5
2. An OB-R isoform according to Claim 1 which is substantially pure.
3. An OB-R isoform according to Claim 1 which is a
10 c' isoform.
4. An OB-R isoform according to Claim 1 which is an f isoform.
- 15 5. An OB-R isoform according to Claim 1 which is a g isoform.
6. An OB-R isoform according to Claim 1 which is from a rat.
20
7. An OB-R isoform according to Claim 6 which is from a wild-type rat.
8. An OB-R isoform according to Claim 6 which is
25 from a *fatty* rat.
9. An OB-R isoform according to Claim 3 which is human.
- 30 10. An OB-R isoform according to Claim 4 which is human.
11. An OB-R isoform according to Claim 5 which is human.

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12. An OB-R isoform according to Claim 3 which is from a mouse.
- 5 13. An OB-R isoform according to Claim 4 which is from a mouse.
14. An OB-R isoform according to Claim 5 which is from a mouse.
- 10 15. A nucleic acid encoding an OB-R of Claim 1.
16. A nucleic acid according to Claim 15 which is a cDNA.
- 15 17. A vector comprising a nucleic acid which encodes an OB-R of Claim 1.
18. A vector according to Claim 17 which is a plasmid.
- 20 19. A host cell containing a vector according to Claim 17.
- 25 20. A host cell according to Claim 19 which is *E. coli*, a mammalian cell, or a yeast cell.
- 30 21. An assay to determine if a putative ligand binds to an OB-R isoform c', f or g comprising: contacting the putative ligand with an OB-R isoform c', f or g, and determining if binding has occurred.

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22. An assay according to Claim 17 wherein the ligand is labeled.
23. An assay to determine if a putative ligand binds to
5 an OB-R isoform c', f or g which is a trans-activation assay.

1 MTCQKFYVVL LHWEFLYVIT ALNLAYPTSP WRFKLFCAPP STTDDSFLLSP
51 AGVPNTSSL KGASEALVEA KFNSTGIYVS ELSKTIFHCC FGNEQGQNC
101 ALTGNTEGKT LASVVKPLVF RQLGVNWIDIE CWMKGDLTLF ICHMEPLLKN
151 PFKNYDSKVK LLYDLPEVID DLPLPPLKDS FQTVQCNCV RECECHVPVP
201 RAKVNYALLM YLEITSAGVS FQSPLMSLQP MLVVKPDPPL GLRMEVTDDG
251 NLKISWDSQT KAPFPLQYQV KYLENSTIVR EAAEIVSDTS LLVDSVLPGS
301 SYEVQVRSKR LDGSGWWSDW SLPQLFTTQD VMYFPPKILT SVGSNASFCC
351 IYKNNENQNTIS SKQIVWWMMNL AEKIPETQYN TVSDHISKVT FSNLKATRPR
401 GKFTYDAVIC CNEQACHHRY AELYVIDVNI NISCETDGYL TKGMTCRWSPS
451 TIQSLVGSTV QLRYHRRSLY CPDNPSIRPT SELKNCVLTQ SELKNCVLTQ DGFYECVFQP
501 IFLLSGYTMW IRINHSSLGSL DSPPTCVLPD SVVKPLPPSN VKAETITNTG
551 LLKVSWEKPV FPENNQFQI RYGLNGKEIQ WKTHEVFDAK SKSASLPPSD

FIG. 1A

601 LCAVYVVQVR CRRLDGLGYW SNWSSSPAYTL VMDVKVPMRG PEFWRIMGD
651 ITKKERNVTL LWKPLMKNDS LCSVRRVVK HRTAHNGTWS QDVGNQTNLT
701 FLWAESAHTV TVLAINSIGA SLVNFNLTFS WPM SKVNAVQ SLSAYPLSS
751 CVILSWTLSP NDYSLLYLVI EWKNLNDDG MKWLRLIPSNV NKYYIHDFI
801 PIEKYQFSLY PVFMEGVGKP KIINGFTKDD IAKQQNDAGL YVIVPIIIS
851 CVLLLCTLLI SHORMKJLFW DDVNPNKNCs WAQCLNFQKP ETFEHLFTKH
901 AESVIFGPLL LEPEPVSEEI SVDTAWNKND EMVPAAMVSL LLTTPDSTRG
951 SICISDQCNS ANFSGAQSTQ GTCEDECQSQ PSVKYATLVS NVKTVETDEE
1001 QGAIHSSVSQ CIARKHSPLR QSFSSNSWEI EAQAFFLSD HPPNVISPQL
1051 SFSGCLDELLE LEGNFPPEENH GEKSVYILGV SSGNKRENDM LLTDEAGVLC
1101 PFPAHCLFSD IRILQESCSH FVENNLNLT SGKNFVPMYMP QFQSCSTHSH
1151 KIIENKOMCDL TV

FIG. 1B

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1 TCGGGCAATT GGGCTGACCT TTCTTAACTT GGGATGTGCC TTGGAGGACT
 51 ATGGCTGTCT ATCTCTGAAG TAAGATGACG TGTCAAGAAAT TCTATGTGGT
 101 TTGTTAACAC TCGGAATTTC TGTATGTAT AACTGCACCT AACCTGGCCT
 151 ATCCAACCTC TCCCTGGAGA TTTAAGCTGT TTGCTGGGCC ACCGAGTACA
 201 ACTGATGACT CCTTTCCTCTC TCCTGCTGGA GTCCCCAAACA ATACTTCGTC
 251 TTTGAAGGGG CCTTCTGAAG CACTTGTGA AGCTAAATT AATTCAACTG
 301 GTATCTACGT TTCTGACTTA TCCAAAAACCA TTTCACCTG TTGCTTTGGG
 351 AATGAGCAAG GTCAAAACCTG CTCCGGACTC ACAGGCAAAACA CTGAAGGGAA
 401 GACGCTGGCT TCAGTGGTGA AGCCTTTAGT TTTCGGCAA CTAGGTGTAA
 451 ACTGGGACAT AGAGTGGCTGG ATGAAAGGGG ACTTGACATT ATTCAATCTG
 501 CATATGGAAC CATTACTTAA GAACCCCTTC AGAAATTATG ACTCTAAAGGT
 551 TCACCTTTA TATGATCTGC CTGAAAGTTAT AGATGATTG CCTCTGCC
 601 CACTGAAAGA CAGCTTCAG ACTGTCCAGT GCAAATGCAAG TGTTCGGAA
 651 TGGCAATGTC ATGTACCACT ACCCAGAGCC AAAGTCAACT ACCGCTCTTC

FIG. 2A

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701 GATGTATTAA GAAATCACAT CTGCTGGTGT GAGTTTCAG TCACCTCTAA
 751 TGTCACTGCA GCCCATGCTT GTTGTGAAGC CCGATCCACC GCTGGGTTTG
 801 CGTATGGAAAG TCACAGATGA TGGTAATTAA AAGATTCAAT GGGACAGCCA
 851 AACAAAGCA CCATTCCAC TTCAATTATCA CCTCAAAATAT TTAGAGAATT
 901 CTACAACTCGT AAGAGGGCT GCTGAAATCG TCTCGGATAAC ATCTCTGCTG
 951 GTAGACAGCG TGCTTCCTGG GTCTTCATAC GAGGTCCAGG TGAGGAGCAA
 1001 GAGACTGGAT GGCTCAGGAG TCTGGAGTGA CTGGAGTTA CCTCAACTCT
 1051 TTACCAACACA AGATGTCAATG TATTTCAC CCAAATTTCT GACGGAGTGT
 1101 GGATCCAATG CTTCCTTTG CTGCATCTAC AAAAATGAGA ACCAGACTAT
 1151 CTCCTCAAAA CAAATAGTTT GGTGGATGAA TCTAGCCGAG AAGATCCCCG
 1201 AGACACAGTA CAAACACTGTC AGTGACCCACA TTAGCAAAGT CACTTTCTCC
 1251 AACCTGAAAG CCACCAAGACC TCGAGGGAAAG TTACCTATG ATGGAGTGTAA
 1301 CTGCTGCAAT GAGCAGGGCAT GCCATCACCG CTACGGCTGAA TTATATGTGA

FIG. 2B

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1351 TCGATGTCAA TATCAATA TCATGTGAAA CTGACCGGTA CTTAACTAAA
 1401 ATGACTTGCA GATGGTCACC CAGCACAAATC CAATCACTAG TGGGAAGGCAC
 1451 TGTGCAGTTG AGGTATCACA GGGCCAGCCT GTACTGTCCC GATAATCCAT
 1501 CTATTCGTCC TACATCAGAG CTCAAAAACT GCGTCTTACA GACAGATGGC
 1551 TTTTATGAAT GTGTTTTCCA GCCAATCTTT CTATTATCTG GCTATACAAT
 1601 GTGGATCAGG ATCAACCATT CTTTAGGTTTC ACTTGACTCT CCACCAACGT
 1651 GTGCTCTTCC TGACTCCGTA GTAAAACCACT TACCTCCATC TAATGTAAAA
 1701 GCAGAGATTAA CTATTAACAC TGGATTATTC AGACTATCTT GGGAAAAGCC
 1751 AGTCCTTCCA GAGAATAACC TTCAAGTCCA GATTGGATAT GGCTTAAATG
 1801 GAAAAGAAAT ACAATGGAAAG ACACACGGG TATTCCGATGC AAAATCAAAA
 1851 TGGGCCAGCC TGCCAGTCTC AGATCTCTGT GCGGTCTATG TGGTACAGGT
 1901 TCGCTGCCGG CGGTGGATC GACTAGGGTA TTGGACTAAT TGGAGGCACTC
 1951 CAGCCTACAC TCTTGTCTATG GATGTAAGAAG TTCCCTATGAG AGGGCCCTGAA
 2001 TTCTGGAGAA TAATGGATGG GGATATTACT AAAAAGGAGA GAAATGTCAC

FIG. 2C

2051 CTTGCTTGG AAGCCACTGA TGAAAATGA CTCACTGTGT AGTGTGAGGA
2101 GGTATGTCGT GAAGCATCGT ACTGCCACAA ATGGACATG GTCACAAAGAT
2151 GTGGAAATC AGACCAATCT CACTTTCTG TGGGAGAAAT CAGCACACAC
2201 TGTACAGTT CTGGCCATCA ATTCCATCGG TGCCTCCCTT GTGAATTATA
2251 ACCTTACGTT CTCACTGGCC ATGAGTAAG TGAATGCTGT GCAGTCACTC
2301 AGTGCTTATC CCCTGAGCAG CAGCTGCCGT C ATCCCTTCCTT GGACACTGTGTC
2351 ACCTAATGAT TATAGTCTGT TATATCTGGT TATTGAATGG AAGAACCTTA
2401 ATGATGATGA TGGATGAG TGGCTTAGAA TCCCTTCGAA TGTTAACAAAG
2451 TATTATATCC ATGATAATT TATTCTTATC GAGAAATATC AGTTAGTCT
2501 TTACCCAGTA TTTATGGAAAG GAGTTGGAAA ACCAAAGATA ATTATGGTT
2551 TCACCAAGA TGATATGCC AAACAGCAA ATGATGCAGG GCTGTATGTC
2601 ATGTACCGA TAATTATTC CTCTTGTCGT CTGGTGTGTC GAACACTGTGTT
2651 AATTTCACAC CAGAGAAATGA AAAACTTGT TGGGAGCAT GTTCCAAACC

FIG. 2D

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2701 CCAAGAATTG TTCTGGCA CAAGGACTTA ATTCCAAA GCCTGAAACA
2751 TTGAGGCATC TTTTACCA GCATGGAGA TCAGTGATAT TTGGTCCCTCT
2801 TCTCTGGAG CCTGAACCAG TTTCAGAAGA AATCAGTGTCA GATACAGCTT
2851 GGAAAATAA AGATGAGATG GTACCAGCAG CTATGGTCTC ACTTCTTTG
2901 ACCACTCCAG ATTCCACAAAG GGGTCTATT TGTATCAGTG ACCAGTGAA
2951 CAGTGTAC TCTCTGGG CTCAGGCCAC CCAGGGAAACC TGTGAGGATG
3001 AGTGTAGAG TCAACCCTCA GTTAAATATG CAACGGCTGGT CAGGAAACGTG
3051 AAAACAGTGG AAACGTGATGA AGAGCAAGGG GCTATAACATA GTTCTGTCA
3101 CCAGTGCATC GCCAGGAAAC ATTCCCCACT GAGACAGTCT TTTTCTAGCA
3151 ACTCCTGGGA GATAGAGGCC CAGGCATTTC TCCTTTTATC AGATCATCCA

FIG. 2E

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3201 CCCAATGTGA TTTCACCACA ACTTTCATTC TCAGGGCTGG ATGAGCTTT
3251 GGAACCTGGAG GGAATTTTC CTGAAAGAAGA TCACGGGGAA AAATCTGTGT
3301 ATTATCTAGG ACTCTCCTCA GGAAACAAAAA GAGAGAATGA TATGCTTTG
3351 ACTGATGAGG CAGGGCTATT GTGCCCATTC CCAGCTCACT GTCTGTTCAG
3401 TGACATCAGA ATCCCTCCAGG AGAGCTGTTC ACACTTGTA GAAAATAATT
3451 TGAATTAGG GACCTCTGGT AAGAACTTTC TACCTTACAT GCCCCACTTT
3501 CAATCCTGTT CCACTCACAG TCATAAGATA ATAGAAAATA AGATGTGTGA
3551 CTTAAACTGTG TAATCTTGTG CAAAAACTTC CAGGTTCCAT TCCAGTAGAG
3601 TGTGTCAATGTT ATAATATGTT CTTTATAGT TGTGGGTGGG AGAGAAAGCC

FIG. 2F

1 TGGGCAATT GGCTGACCT TTCTTATGCT GGGATGTGCC TTGGAGGACT
51 ATGGGTGTCT ATCTCTGAAG TAAGATGACG TGTCAAGAAT TCTATGTGGT
101 TTGTTACAC TGGGAATTTC TGTATGTGAT AACTGCACTT AACCTGGCCT
151 ATCCAACCTC TCCCTGGAGA TTTAAGCTGT TTTGTCGCC ACCGAGTACA
201 ACTGATGACT CCTTTCTCTC TCCTGCTGGA GTCCCCAACAA ATACTTCGTC
251 TTGAAAGGG GCTTCTGAAG CACTTGTGA AGCTAAATT ATTCAACTG
301 GTATCTACGT TTCTGAGTTA TCCAAAACCA TTTTCCACTG TTGCTTGGG
351 AATGAGCAAG GTCAAAACTG CTCCGCACTC ACAGGCAACAA CTGAAGGGAA
401 GACGCTGGCT TCAGTGGTGA AGCCCTTAGT TTTCCGCCAA CTAGGTGAA
451 ACTGGGACAT AGAGTGTGG ATGAAAGGG ACTTGACATT ATTCACTCTG
501 CATATGGAAC CATTACTAA GAACCCCTTC AAGAATTATG ACTCTAAGGT
551 TCACCTTTA TATGATCTGC CTGAAGTTAT AGATGATTG CCTCTGCC
601 CACTGAAAGA CAGCTTTCAAG ACTGTCCAGT GCAACTGCAG TTGTCGGGAA

FIG. 3A

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651 TCGGAATGTC ATGTACAGT ACCCAGAGCC AAAGTCAACT ACGCTCTTCT
701 GATGTATTAA GAAATCACAT CTGCTGGTGT GAGTTTCAG TCACCTCTAA
751 TGTCACTGCA GCCCATGCTT GTTGTGAAGC CCGATCCACC GCTGGGTTTG
801 CGTATGGAAG TCACAGATGA TGGTAATTAA AAGATTTCAT GGGACAGCCA
851 AACAAAGCA CCATTTCAC TTCAATATCA GGTGAATAT TTAGAGAAATT
901 CTACAATCGT AAGAGAGGCT GCTGAAATCG TCTCGGATAC ATCTCTGCTG
951 GTAGACAGCG TGCTTCCTGG GTCTTCATAC GAGGTCCAGG TGAGGAGCAA
1001 GAGACTGGAT GGCTCAGGAG TCTGGAGTGA CTGGAGTTA CCTCAACTCT
1051 TTACCAACACA AGATGTCAATG TATTTCAC CCAAATTCT GACGGAGTGT
1101 GGATCCAATG CTTCCCTTTC CTGCATCTAC AAAATGAGA ACCAGACTAT
1151 CTCCTCAAAA CAAATAGTTT GGTGGATGAA TCTAGCCGAG AAGATCCCG
1201 AGACACAGTA CAACACTGTC AGTGACCACA TTAGCAAAGT CACTTCTCC
1251 AACCTGAAAG CCACCAAGACC TCGAGGGAAG TTACCTATG ATGCAGTGTAA

FIG. 3B

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1301 CTCGCTGCAAT GAGCAGGGCAT GCCCATCACCCG CTACGGCTGAA TTATATGTGA
1351 TCGATGTCAA TATCAAATA TCATGTGAAA CTGACGGGTA CTTAACTAAA
1401 ATGACTTGTCA GATGGTCACC CAGGCACAAATC CAATCACTAG TGGGAAGGCAC
1451 TGTGCAGTGT AGGTATCACCA GGCGCAGCCT GTACTGTCCC GATAATCCAT
1501 CTATTCGTCC TACATCAGAG CTCAAAACACT GCGTCTTACA GACAGATGGC
1551 TTTTATGAAT GTGTTTCCA GCCAATCTTT CTATTATCTG GCTATACAACT
1601 GTGGATCAGG ATCAACCATT CTTTAGGTTTC ACTTGACTCT CCACCAACGT
1651 GTGTCTTCC TGACTCCGTA GTAAAACCAC TACCTCCATC TAATGTAAAA
1701 GCAGAGATTA CTATAAACAC TGGATTATTG AAAGTATCTT GGGAAAAGCC
1751 AGTCTTTCCA GAGAATAACC TTCAGTTCCA GATTGGATAT GGCTTAATG
1801 GAAAAGAAAT ACAATGGAAG ACACACGAGG TATTGGATGC AAAATCAAAA
1851 TCGGCCAGGCC TGCCAGTGT AGATCTCTGT GCGGTCATG TGGTACAGGT

FIG. 3C

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1901 TCGCTGCCGG CGGTGGATG GACTAGGGTA TGGAGTAAT TGGAGCAGTC
 1951 CAGCCTACAC TCTTGTCAATG GATGTAAGAAG TCCCTATGAG AGGGCCTGAA
 2001 TTCTGGAGAA TAATGGATGG GGATATTACT AAAAAGGAGA GAAATGTCAC
 2051 CTTGCTTTGG AAGCCACTGA TGAAAATGA CTCACGTGT AGTGTGAGGA
 2101 GGTATGGGT GAAGCATCGT ACTGCCACCA ATGGGACATG GTCACAAAGAT
 2151 GTGGAAATC AGACCAATCT CACTTTCTG TGGGCAGAAAT CAGCACACAC
 2201 TGTTACAGTT CTGGCCATCA ATTCCATCGG TGCCTCCCTT GTGAATTTTA
 2251 ACCTTACGTT CTCATGGCCC ATGAGTAAG TGAAATGCTGT GCAGTCACTC
 2301 AGTGCTTATC CCCTGAGCAG CAGCTGCGTC ATCCTTCCCT GGACACTGTC
 2351 ACCTAATGAT TATAGTCTGTT TATATCTGTT TATTGAATGG AAGAACCTTA
 2401 ATGATGATGA TGGAAATGAAG TGGCTTAGAA TCCCTTCGAA TGTTAACAAAG
 2451 TATTATATCC ATGATAATT TATTCCATC GAGAAATATC AGTTTAGCTCT

FIG. 3D

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2501 TTACCCAGTA TTTATGGAAAG GAGTTGGAAA ACCAAAGATA ATTAATGGTT
2551 TCACCAAAGA TGATATCGCC AACAGCAA ATGATGCAGG GCTGTATGTC
2601 ATTGTACCGA TAATTATTTC CTCTTGTGTC CTGCTGCTCG GAACACTGTT
2651 ATTTCACAC CAGAGAATGA AAAAGTTGTT TTGGGACGAT GTTCCAAACC
2701 CCAAGAATTG TTCCCTGGCA CAAGGACTTA ATTCCAAAA GATAATGCCTG
2751 GCAGAAATTG GAGGATATAG AGTGGATGCC GTCAAATGCC TTTAGACTCT
2801 GGCTTCCCTG GCTGTCTCAC ATCTCCCTA TTGGAGCTAA GTGTGGTGT
2851 GTATTAGCA GGGTATCTGG CAGATATTG CAGATATTG AAATAATCACC
2901 CTAAAATTCC AGATTCTGGT AAACCTGAAGT GAATTTCAGA AATTATTGTA
2951 TTTATGTGT TGCACATATG TGTGCAGGTA CCCACCGAAA TCTGCAGAGG
3001 CATCAGATGC CCCAGAGCTG GAACTGACAG TTGTGACCCCT GATATGAGTT
3051 CTGGGAATGA GCTCAGTCCT CTGGAAAGAGC TGCAAGCACT ATTAACGTGCT

FIG. 3E

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3101 GAGCCATCTT TTCAGTCCCT CATGTATAGA TTAAAAAAA TTGGGGTTTG
3151 AAGATCCTCA TTGTGAGAA ATTCCCTCTT ACCTTTGCAA ACACCTTTTC
3201 TCATTTTAG TATATGTATT CATATTTCAC TGTCTCATT TCATATATG
3251 TGGTCACAGT TTTAAGTAT TTCTAAGGCA TAACAAAGAT GTAAATATTAA
3301 GAATAATAA AAGAATAAT CATAATCCA GATGGTAGTG ACAGACACCT
3351 TTAATCCCAG TACTAAGGAG ACAGAGATAG GTAAATCTGT ATGAAATTGAG
3401 GACACGCCCTG TTCTACAAAG AAATTCAGG ACATCTAGGG GTATCCACAA
3451 AGAAACACTG TCTCAAAAAA TGCCAAACAA TCAAAAAAA AAAA

FIG. 3F

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1 GTCACTTTT AAGTATTAC CCAAGATATC TAAGGTTGCA GTTTAGATAC
51 TCTATTACAT AGAGATCTTT AACACATCTTT AAAAGGCTTT ATTGTTGTCCT
101 GTTCACTTA TTAATCCCGT TTATCCTTTG TCTATAGCAA TAGCTGGTT
151 TTGGATTGTA TCAGAGGAAA CAAAGTTCAAG TCATTATCA CATGAGAGTT
201 GACAAGGTTGT CTTTTTTTT TCTCGTCACT GTACATAAAA AAATAAATAC
251 TACAAGAGGA AGGAACATTG TAGATGGAGA ATAGATAACT GACTAAAAGG
301 GCTTTCTTTA GTCAAAAAGT TTAGGATCAA TATTATGAGT TTCTGATATT
351 CAATATTCA CCATGACTTA CAAGTACAGT GTTGTTTT

FIG. 4

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US97/07521

A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) :Please See Extra Sheet.
US CL :Please See Extra Sheet.

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 435/69.1, 69.5, 252.3, 320.1, 7.1, 7.2; 536/23.5; 530/350, 351; 514/2.8,12

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)
APS, BIOSIS, CA

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	TARTAGLIA et al. Identification and expression cloning of a Leptin Receptor, OB-R. Cell. 29 December 1995, Vol. 83, pages 1263-1271, see entire document.	1-5, 9-20 -----
Y	CHUA et al. Phenotypes of Mouse <i>diabetes</i> and Rat <i>fatty</i> due to mutations in the OB (Leptin) Receptor. Science. 16 February 1996, Vol. 271, see pages 994-996.	1-23 -----
X	CHEN et al. Evidence that the diabetes gene encodes the Leptin Receptor: Identification of a mutation in the Leptin Receptor gene in <i>db/db</i> mice. Cell. 09 February 1996, Vol. 84, pages 491-495, see entire document.	1-8, 12-20 -----
Y		1-23 -----

 Further documents are listed in the continuation of Box C. See patent family annex.

* Special categories of cited documents	T	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
*'A' document defining the general state of the art which is not considered to be of particular relevance		
*'E' earlier document published on or after the international filing date	'X'	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
*'L' documents which may throw doubt on priority claim(s) or which is cited to establish the publication date of another citation or other special reasons (as specified)	'Y'	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
*'O' document referring to an oral disclosure, use, exhibition or other means	'A'	document member of the same patent family
*'P' document published prior to the international filing date but later than the priority date claimed		

Date of the actual completion of the international search

19 JUNE 1997

Date of mailing of the international search report

7 JUL 1997

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Form PCT/ISA/210 (second sheet)(July 1992)*

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US97/07521

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	CIOFFI et al. Novel B219/OB receptor isoforms: Possible role of leptin in hematopoiesis and reproduction. <i>Nature Medicine</i> . May 1996, Vol. 2, No. 5, pages 585-589, see entire document.	1-5, 9-20 -----
Y		1-23
X	WO 96/08510 A1 (PROGENITOR, INC.) 21 March 1996 (21.03.96), see the figures and claims.	1-23
X	LEE et al. Abnormal splicing of the leptin receptor in <i>diabetic</i> mice. <i>Nature</i> . 15 February 1996, Vol. 379, pages 632-635, see entire document.	1-5, 12-20 -----
Y		1-23
X	HODGSON J. Receptor screening and the search for new pharmaceuticals. <i>Bio/Technology</i> . September 1992, Vol. 10, pages 973-997, see entire document.	21-23
X	CA 2,104,996 A1 (BEHRINGWERKE AKTIENGESELLSCHAFT) 01 March 1994 (01.03.94), see the claims.	21-23

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US97/07521

A. CLASSIFICATION OF SUBJECT MATTER:
IPC (6):

C12P 21/00; C12N 1/20, 15/00; G01N 33/53; C07H 21/04; C07K 1/00, 14/52; A61K 45/05, 38/19, 38/00

A. CLASSIFICATION OF SUBJECT MATTER:
US CL :

435/69.1, 69.5, 252.3, 320.1, 7.1, 7.2; 536/23.5; 530/350, 351; 514/2,8,12